

Phytophthora Root Rot Resistance

Phytophthora medicaginis Hansen & Maxwell, sp. nov.
 Judy A. Thies and Donald K. Barnes

PLANT CULTURE

Field Methods(3)

No. of Plants 40 to 60 per replication
 No. of Reps 3 or 4 replications
 Other Plant in early May on a level area with relatively heavy soil, but with good internal drainage

Greenhouse Method

Container 20 cm deep watertight tanks with drain holes and a 2.5 cm diameter pipe placed vertically in one corner to allow flooding sand from bottom of tank
 Medium Washed, pasteurized sand
 Temp/Light 20 to 24°C (sand temperature); 16 hour daylength
 No. of Plants 25 per replication
 No. of Reps 4 minimum
 Other Inoculate with *Rhizobium meliloti* Dang and fertilize

INOCULUM CULTURE

Greenhouse Method

Source Culture on V-8 juice agar in 9 cm petri dishes
 Maintenance Store cultures on V-8 juice agar at 4°C

INOCULATION PROCEDURE

Field Method

Type of Inoc Select a field that is naturally infested; or spread infested soil from several geographic areas over the field, incorporate to a depth of about 15 cm and grow a susceptible variety for one year prior to using the area

Greenhouse Method

Age of Plant Plant seed into sand
 Type of Inoc Two week old cultures
 Concentration Mix inoculum with sand before planting at a rate of one petri dish per 500 cm² surface area

INCUBATION

Field Method

Plant Counts Count plants (alive + dead) when seedlings are in the unifoliolate stage
 Culture About 4 weeks after planting, irrigate each day for 3 weeks to keep soil continuously saturated, allow soil to dry for 1 week, clip plants, and cultivate soil. Repeat the sequence two more times. Spray for insects as needed
 Row Spacing Approximately 0.3 m
 Age at Rating 14 to 15 weeks after planting

Greenhouse Method

Plant Counts Same as field method
 Culture Water seedlings sparingly until they are well established (4 weeks), plug drain holes and water daily to raise water level surface; maintain flooded conditions for about 4 weeks
 Row Spacing Approximately 3.5 cm
 Age at Rating 8 weeks after planting

RATING

Dig all plants retaining 25 cm or more of the taproot. Spray roots to remove excess soil, bundle plants, and soak roots in a tub of water. Complete washing and rate plants indoors under uniform light.

- 1 Resistant Roots clean no lesions; many small rootlets on taproot
- 2 Resistant Small root lesions (2mm); small rootlets absent
- 3 Susceptible Large nongirdling root lesion(s) and/or branch root tips rotted off
- 4 Susceptible Extensive lesions with ends of large tap or lateral roots rotted off 10 cm or more below the crown.
- 5 Susceptible Tap and lateral roots almost destroyed; plant alive.
- 6 Susceptible Plants dead (calculated as loss in stand)

CHECK CULTIVARS

	Approximate Expected Reaction (%)	Acceptable Range of Resistance (%)
Resistant		
Agate**	43	25-55
Susceptible		
Saranac**	3	0-10

Values for resistant standards include total of 1's and 2's.

DISTRIBUTION AND SEVERITY OF PHYTOPHTHORA ROOT ROT



Phytophthora root rot, *Phytophthora megasperma* Drechs. f. sp. *medicaginis* (3)

Click on the map above for a larger version. See also the [KEY](#).

SOURCES OF INOCULUM AND EXPERTISE

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CORRELATION TO FIELD REACTION

Field and greenhouse evaluations were correlated ($r = 0.99$ and 0.95) in two tests. (3). Field tests tend to be more precise, with greenhouse tests useful for screening.

PATHOTYPES

Isolates of *Phytophthora megasperma* with different levels of pathogenicity on alfalfa cultivars exist (2). Therefore, it is important to use a mixture of highly pathogenic isolates.

PLANT GROWTH OPTIONS AND RANGE OF CONDITIONS

Monitor root rot development during the season. Symptoms can be increased or reduced by changing the frequency and/ or amount of irrigation. Potato leafhopper control is very important in the Midwest.

HELPFUL INFORMATION

Ratings may be expressed as an average severity index (A.S.I.) which is most precise, or percentage of resistant plants (3) which can be adjusted to a standard check to compare entries between tests. The percentage of plants adjusted to Agate is useful for comparing cultivars tested in different years.

ALTERNATIVE METHODS

Greenhouse tests using zoospores (1) in a method analogous to that used for *Aphanomyces* (4) has been successfully used to rank lines although the percent resistant plants is somewhat lower than field test results.

REFERENCES

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